

Importance of Epigenetic Testing in Breast Cancer Patients [Version 1, Awaiting Peer Review]

Eva Jezkova and Marian Adamkov*

Department of Histology and Embryology, Jessenius Faculty of Medicine, Comenius University in Bratislava, Slovakia

***Corresponding author:** Marian Adamkov, Department of Histology and Embryology, Jessenius Faculty of Medicine, Comenius University in Bratislava, Mala Hora 4, 036 01 Martin, Slovakia, Email: Marian.Adamkov@jfmed.uniba.sk

Copyright: © 2017 Eva Jezkova and Marian Adamkov. This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source.

Original Submission

Received: June 30, 2017

Accepted: July 12, 2017

Published: July 19, 2017

Open Peer Review Status: Awaiting Peer Review

How to cite this article: Eva Jezkova, Marian Adamkov. Importance of Epigenetic Testing in Breast Cancer Patients [Version 1, Awaiting Peer Review]. Insights Breast Cancer (2017) 1: 4.1

Insights in Breast Cancer

Introduction

Frequency of breast cancer is increasing worldwide and is the most common cancer in women in most countries. The screening of mammary carcinoma is performed in the female population nationwide anywhere in the world. However, we have no available screening methods that would be effective and would meet the criteria of simplicity, non-invasiveness and financial difficulty. As is well known, the deregulation of epigenetic mechanisms in conjunction with genetic changes causes some genetic diseases, including cancer. Epigenetic mechanisms are spontaneous and reversible, they represent differences in DNA methylation and in the structure of chromatin. The number and pattern of methylated cytosines affects the function of the gene, low methylation leads to high activity, and vice versa. It plays an essential role during the process of differentiation and development of vertebrates, gene imprinting and X chromosome inactivation. In DNA isolated from malignant tumor cells, abnormal hypermethylation of CpG islands was observed in the promoter regions of genes, one of the major regulatory regions of the gene. In malignant tumors, transcription of tumor suppressor genes is also suppressed as a result of DNA methylation, and this affects cell cycle regulation. One of the options for targeting cancer treatment is to regain the gene expression by DNA-inhibition of methyltransferases. The constant position of abnormal CpG islands methylation in the tumor suppressor gene promoter region allows for a simpler detection strategy as is possible for many common mutations in cancer. This implies that the same primer combination could be used for all patients to detect tumor-specific methylation changes of the gene by means of a methylation-sensitive, high-resolution analysis of the melting curves. These promoter hypermethylation characteristics, along with the fact that all tumors have 1 or more hypermethylated loci, make these changes valuable as DNA markers for sensitive and early tumor detection. Thanks to the knowledge gained from gene expression regulation studies, it is possible to identify the specific causes of the disease and to develop therapeutic strategies on the basis of this, which could lead to the restoration of the normal epigenetic state [1-3].

Materials and Methods

Immunohistopathological classification including the tumor (pT), degree of differentiation (G), lymph node (pN) status, estrogen (ER) and progesterone (PR) receptor statuses, and HER2 gene amplification were determined by a qualified pathologist. The tumor type was evaluated on the basis of the WHO classification of tumors [4,5], the histological stage, and the classification was performed in accordance with the revised pTNM classification of AJCC and UICC [6]. The estrogen and progesterone receptor status was expressed as a percentage of positive reactive cells from the total number of captured cells. A positive ER or PR state was considered to be a case in which $\geq 1\%$ of the cells were positive [7]. The HER2 amplification status was detected by immunocytochemistry (HerceptTest) and eval-

uated according to the criteria currently published by ACSO / CAP 2007 [8]. DNA for the purpose of the methylation analysis was subjected to deparaffinization from the paraffin sections of the tissue in the tube. The DNA extraction itself was performed by using DNeasy Blood & Tissue Kit (Qiagen, Germany), where we followed the manufacturer's recommendations. The method of the methylation-specific PCR (MSP) was performed on a MiniOpticon PCR Real-Time PCR system (Bio-Rad, Applied Biosystems, California, USA). For MSP, internal primers were used for both the methylated and non-methylated sequence of the *RASSF1A* gene. The MSP analysis allowed detection of the presence of methylation in genomic DNA by comparison with the commercially available methylated and unmethylated DNA standards (EpiTect, Qiagen, Germany). All statistical tests were performed using R software [9]. Statistically significant P values were equal to or less than 0.05 ($P \leq 0.05$)

Results

We found the association of the methylation status of the *RASSF1A* gene with histopathological parameters. The association of the methylation status of the *RASSF1A* gene with the tumor grade (G) was significant in all grades ($P < 0.0001$ for G1, G2 and G3). Significant association was observed between stage 3 of ductal breast cancer and the positive methylation status of the *RASSF1A* gene ($p < 0.05$). The metastatic status of the tumor stage (pT) was significantly associated with the methylation of *RASSF1A* gene ($P < 0.0001$ for pT1 and pT2, $P < 0.05$ for pT4). The relationship between the level of methylation and the type of breast cancer was strongly associated only in the ductal invasive carcinoma (DIC) ($P < 0.0001$) samples for the *RASSF1A* gene. Presence or absence of methylation in *RASSF1A* promoter region was investigated by MSP in 116 samples of primary breast tumors, where we found out methylated alleles in 97 (83,6%) samples.



Observation 1: Result from electrophoresis in agarose gel shows representative results from MSP method. The presence of MSP products in lines 3,4,6,7,9,10,11,12,13,14 confirms presence of methylated alleles in promoter region of *RASSF1A* gene. The absence of MSP products in lines 5 and 8 confirms presence of unmethylated alleles in *RASSF1A* gene. Line 1 is methylated DNA standard and in line 2 is unmethylated DNA standard. Line "M" shows 100bp marker, and line 15 is negative control. Product size of *RASSF1A* gene is 198 bp.

Discussion

Recent studies show the diagnostic and prognostic potential of *RASSF1A* methylation. In relation to breast cancer, the incidence of *RASSF1A* methylation occurs in many independent studies in varying degrees. Honorio et al [10] demonstrated the presence of the *RASSF1A* promoter methylation in 65% invasive

Insights in Breast Cancer

breast cancer and 42% of the relevant ductal carcinoma in situ (DCSI). Burbee et al [11] recording 49% primary breast tumors with methylated *RASSF1A*. The largest occurrence of methylation of the *RASSF1A* promoter (95%) in breast cancer tissue was recorded by Yeo et al [12]. A recent study found a link between ER-positive and ER-negative status in relation to epigenetic changes in genes associated with breast cancer, including the *RASSF1A* gene [13].

Conclusion

Breast cancer is a heterogeneous disease with very different responses to therapy and varying lengths of survival. Overall is determined by histopathological criteria based on the size, level of invasiveness and infiltration of lymph nodes, and on the immunohistochemical characteristics of cell surface receptors, including estrogen receptor (ER), progesterone receptor (PR) and human epidermal growth factor receptor 2 (HER2). In many cases, however, determining the degree of breast cancer is not sufficient to predict prognosis or response to treatment for the enormous heterogeneity of the disease. Therefore, new classifications approaches based on epigenetic patterns are now emerging. DNA methylation is a strong epigenetic biomarker, significantly more stable than RNA or proteins, and is therefore a promising target for developing new approaches to determining the diagnosis and prognosis of breast and other tumors. The aim of this study was to evaluate the presence of methylation of the *RASSF1A* promoter region and its potential relationship with the immunohistopathological characteristics of breast cancer. Our results suggest that hypermethylated gene loci may exhibit a methylated state during carcinogenesis, and in relation to the histopathological parameters, methylation appears to be an early tumor transformation event.

References

1. Dammann R, Yang G, Pfeifer GP. Hypermethylation of the CpG island of Ras association domain family 1A (*RASSF1A*), a putative tumor suppressor gene from the 3p21. 3 locus, occurs in a large percentage of a human breast cancers. *Cancer Res.* 2001; 61: 3105-3109.
2. Vaissière T, Hung RJ, Zaridze D, Moukeria A, Cuenin C, et al. Quantitative analysis of DNA methylation profiles in lung cancer identifies aberrant DNA methylation of specific genes and its association with gender and cancer risk factors. *Cancer Res.* 2009; 69: 243-252.
3. Pfeifer P, Dammann R. Methylation of the tumor suppressor gene *RASSF1A* in human tumors. *Biochemistry.* 2005; 70: 576-583.
4. Lakhani SR, Ellis IO, Schnitt SJ, Tan PH, van de Vijver MJ. *WHO Classification of Tumours of the Breast*, 4th edn. Lyon: IARC. 2012.
5. Elston CW, Ellis IO. *Pathological Prognostic Factors in Breast Cancer*. I. The Value of Histological Grade in

Breast Cancer: Experience from a large study with Long-Term Follow-up. *Histopathology.* 1991; 19: 403-410.

6. Edge SB, Compton CC. The American Joint Committee on Cancer: the 7th edition of the AJCC cancer staging manual and the future of TNM. *Ann Surg Oncol.* 2010; 17: 1471-1474.
7. Hammond MEH, Haves DF, Wolff AC, Mangu PB, Te-min S. American Society of Clinical Oncology/ College of American Pathologists guideline recommendations for immunohistochemical testing of estrogen and progesterone receptors in breast cancer. *Arch pathol Lab Med.* 2010; 134: E1-E16.
8. Wolf AC, Hammond ME, Hicks DG, Dowsett M, McShane LM, et al. American Society of Clinical Oncology; College of American Pathologists. Recommendations for human epidermal growth factor receptor 2 testing in breast cancer: American Society of Clinical Oncology/ College of American Pathologists clinical practice guideline update. *Arch Pathol Lab Med.* 2014; 138: 241-256.
9. R Development Core Team. A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. 2015. <http://www.R-project.org>.
10. Honorio S, Agathangelou A, Schuermann M, Pankow W, Viacava P, et al. Detection of *RASSF1A* aberrant promoter hypermethylation in sputum from chronic smokers and ductal carcinoma in situ from breast cancer patients. *Oncogene.* 2003; 22: 147-150.
11. Burbee DG, Forgacs E, Shivakumar I, Fong K, Gao B, et al. Epigenetic inactivation of *RASSF1A* in lung and breast cancers and malignant phenotype suppression. *J Natl Cancer Inst.* 2001; 93: 691-699.
12. Yeo W, Wong W, Wong N, Law BK, Tse GM, et al. High frequency of promoter hypermethylation of *rassf1a* in tumorous and non-tumorous tissue of breast cancer. *Pathology.* 2005; 37: 125-130.
13. Sunami E, Shinozaki M, Sim MS, Nguyen SL, Vu AT, et al. Estrogen receptor and HER2/neu status affect epigenetic differences of tumor related genes in primary breast tumors. *Breast Cancer Res.* 2008; 10: R46.